

Sleep Medicine 3 (2002) S43-S49

SLEEP MEDICINE

www.elsevier.com/locate/sleep

Modulation of normal and pathologic motoneuron activity during sleep: insights from the neurology clinic, Parkinson's disease, and comments on parkinsonian-related sleepiness

D.B. Rye*

Department of Neurology, Emory University School of Medicine, 1639 Pierce Drive, WMRB-Suite 6000, Atlanta, GA 0322, USA

Keywords: Restless legs syndrome; Periodic limb movement; REM sleep; Nigrostriatal dopamine pathway; Diencephalic dopamine pathway; Monoamine

1. Introduction

A description of electromyographic (EMG) activity during sleep demands familiarity with the organizational principles-anatomical, neurochemical, and physiological-that govern somatic motor system function. The final common pathway to initiate or suppress movements is the lower motor neuron, whose axons innervate striated muscle fibers. The neural circuits governing motoneuron activity are numerous, complex, and for the most part incompletely understood. It is important to speak conceptually of elements that have a significant impact upon motor behavior as 'pre-motor'. Pre-motor neurons include those whose axons directly contact motoneurons and others that, despite being several synapses removed from the neuromuscular junction, elicit movement when stimulated. The precise multisynaptic circuits through which these physiologically defined pre-motor influences reach motoneurons remain illdefined. Pre-motor elements thus describe links of a chain that share in common an ability to modulate motoneuron excitability and thereby movement. This being said, it should become obvious from the discussion that follows that there are many pathways that converge in parallel upon motoneurons, rather than a strictly hierarchical chain built upon serial circuits.

In addition to pre-motor neurons and interneurons intrinsic to the spinal cord, the elements most relevant to this discussion are extrinsic: i.e. they originate from groups of neurons within the brain and are thus described as 'supraspinal' [1]. Current heuristic models of motor system structure and function recognize two divisions: medial and lateral somatic motor systems [2]. The medial system is devoted to modulation of eye, neck, axial, and proximal body move-

* Tel.: +1-404-727-3928; fax: +1-404-727-3157.

E-mail address: drye@emory.edu (D.B. Rye).

ments and includes tectospinal (originating from deep layers of the superior colliculus), vestibulospinal, and reticulospinal (originating from the dorsal two-thirds of the medial tegmentum of the caudal pons and medulla) components. Descending pathways of this system course medially in the spinal cord and terminate principally in the ipsilateral, medial part of the ventral horn containing interneurons and motoneurons of axial and proximal muscles. This system is principally concerned with orienting the head and body relative to the visual fields under control of extrinsic eye muscles. The lateral system is devoted to voluntary movements of the extremities and includes the rubro- and corticospinal tracts. The rubrospinal tract is a phylogenetically old supraspinal system. It is exclusively crossed, descends in the dorsolateral funiculus of the spinal cord, and innervates premotor interneurons, although direct projections to motoneurons also exist. Corticospinal pathways mimic the rubrospinal pathway [3-5], but in parallel with an expanding neocortex, they take on greater specialization and importance in primates [1]. In their descent from the neocortex, many individual corticospinal axons give off collaterals to rubro- and reticulospinal neurons. The majority continue their descent and act monosynaptically upon motoneurons of distal musculature. The primary and supplementary cortices from which the corticospinal tract arise, as well as the red nucleus (the origin of the rubrospinal tract), are both microexcitable; when stimulated, they elicit movements primarily in distal, flexor muscles.

A substantial additional set of pathways originating from the brainstem influence spinal motoneurons. These include monoamine pathways (e.g. dopaminergic, noradrenergic, and serotonergic) and reticulospinal projections originating from the ventromedial medulla [6]. The caudal pons and contiguous ventromedial medulla are remarkable in having neurons that innervate a wide variety of motoneurons, from those innervating distal extremities to diaphragmatic [7],

^{1389-9457/02/\$ -} see front matter © 2002 Elsevier Science B.V. All rights reserved. PII: S1389-9457(02)00148-X

rectus abdominis [8], and pudendal [9] motoneurons. These supraspinal systems have received particular attention because they receive inputs from many forebrain structures designated as 'limbic' in nature or demonstrate activity that is highly correlated with sleep-wake state. These features have justified their consideration as a distinct 'emotional motor system' to many investigators in the field [6,10,11]. The practical clinical significance of these systems lies in the fact that: (1) many commonly prescribed, centrally acting drugs act to enhance or dampen monoaminergic neurotransmission; (2) there are age- and disease-related declines in the densities of many monoaminergic pathways; and (3) the ventromedial medulla also appears to participate in a multisynaptic pathway linking movement-related structures of the basal ganglia with lower motoneurons.

The distributions of the spinal monoamines are very similar to innervation of sensory (dorsal horn), autonomic (intermediolateral cell column), and motor (ventral horn) areas of the spinal cord. Dopaminergic fibers originate predominantly from the diencephalic A11 cell group, and to a lesser extent from the A13 group [12-15]. Spinal noradrenergic innervation originates from the locus coeruleus and subcoeruleal area [16–19]. The serotonergic raphe magnus projects to the dorsal horn, whereas the more caudally situated raphe pallidus and obscurus project principally to the intermediate zone, including direct innervation of Renshaw cells [20] and motoneuron cell groups [21,22]. Glutamate also appears to exist as a cotransmitter in many individual monoaminergic cells [2,23-26]. Two important inhibitory reticulospinal projection systems originating from the ventromedial medulla include pathways containing gamma-aminobutyric acid (GABA) or glycine. Nearly 40% of these appear to be GABAergic, 15% glycinergic, and the remainder a collage of glutamatergic and monoaminergic [11]. That 50% of the terminals descending from the ventromedial medulla and adjacent raphe cell groups make synaptic contact with motoneurons, primarily upon their proximal dendrites [11], emphasizes their relative importance in determining the state of motoneuron excitability. Neural collections in the caudal medulla, dorsal to the inferior olive, seem to exert an inhibitory influence upon motoneurons (e.g. the 'bulbospinal inhibitory zone' of Magoun and Rhines [27]), while rostrally situated neurons at the pontomedullary junction in proximity of the raphe magnus tend to enhance motoneuron excitability [28].

The physiological effects of the spinal monoamine systems are best categorized as 'neuromodulatory'. That is, rather than eliciting excitatory postsynaptic potentials (EPSPs) in motoneurons as does glutamate, or inhibitory postsynaptic potentials in motoneurons as do GABA and glycine, monoamines generally enhance motoneuron responses to excitatory and inhibitory influences [29,30]. By changing the strength of specific synapses and by altering neural membrane properties and specific ion conductances, monoamines rapidly reconfigure motor networks commensurate with environmental and homeostatic demands. By modulating the responsiveness of motoneurons and pre-motor elements to other excitatory or inhibitory inputs, neuromodulators affect the intensity, duration, and timing of motor output [31–33]. Moreover, because individual axons frequently traverse several spinal cord segments, coordination of movement sequences by muscles whose motoneurons are spatially separated (e.g. arm versus leg, proximal versus distal, agonists versus antagonists) is possible.

Long descending monoaminergic fiber systems also appear to have a significant role in modulating ascending sensory transmission either by acting postsynaptically on 'secondary' sensory spinal neurons, or by acting presynaptically upon the terminal axons of dorsal root afferents. Serotonin, noradrenaline, and dopamine, for example, all depress transmission from group II muscle spindle afferents [34–36] and depress motoneuron EPSPs via effects on metabotropic glutamate receptors at both pre- and postsynaptic sites [37]. Thus, prevailing views of monoamine modulation of spinal cord function include shared roles in promoting motor output and inhibiting sensory input.

2. Pathological motoneuron activity in sleep

2.1. Periodic leg movements in sleep (PLMS)

The final common pathway mediating PLMS are neural elements intrinsic to the spinal cord, given their unveiling below the level of pontine infarction [38] or spinal cord transection or pathology [39–42]. A strict derivation from a spinal locomotor network, common to many vertebrates, is unlikely because leg movements detected by standard surface electrodes placed over both anterior tibialis muscles most often occur simultaneously (versus alternately), can exhibit contraction with antagonists (namely, the gastrocnemius [41], personal observations), and may coincide with changes in the activity of upper limb or bulbar musculature (personal observations).

Identification of a single neurophysiological mechanism underlying PLMS remains enigmatic, although the principal deficit manifests as brainstem [43,44] and spinal reflex 'hyperexcitability' [45]. The origin of this enhancement of motoneuron output may reside in any one of several neural substrates outlined above. The recent delineation of statedependent changes in spinal cord excitability demonstrated in restless legs syndrome (RLS)/PLM are of fundamental importance [45]. These changes manifest as: (1) decreased threshold of the flexor reflex (FR); and (2) segmental spread of the FR (from distal to proximal muscles). The FR is a widely studied phenomenon that can be modified by muscle and cutaneous afferents, Renshaw cells, presynaptic inhibition of afferents by intraspinal interneurons, and multiple supraspinal pathways. The pathophysiology of RLS/PLM does not reside in the principal sensory and motor elements themselves based upon observations that waking EMG

S45

activity, resting motoneuron excitability, simple reflexes, and sensory-evoked potentials are generally normal [43,46]. The most powerful and consistent influences impacting upon RLS/PLM originate from outside the spinal cord in supraspinal, pre-motor circuits. This is best exemplified in the setting of spinal cord injury, where pharmacologic agents effective in treating RLS/PLM and dampening FR responses via local spinal circuits (e.g. dopaminomimetics and opioids), are generally, but not universally, *ineffective* [41,42]. This argues that the primary benefit is mediated predominantly by alternate dopaminergic or dopamine-sensitive pathways located supraspinally.

Imaging studies, clinical observations, and animal studies have focused principally upon the nigrostriatal component of the mesotelencephalic dopamine system as a potential key substrate in the pathogenesis of RLS/PLM. Reductions in dopamine uptake into presynaptic axons via the dopamine transporter [47] and in D_2 receptor binding seen in some imaging studies [47,48] suggest a relative excess of extracellular dopamine. However, these changes are of small magnitude and have not been confirmed in a more recent, larger, and better-controlled study [49]. Periodic limb movements also occur when the striatum is depleted of dopamine axons either experimentally [50], or in the face of neurodegeneration occurring with Parkinson's disease (PD) [51,52]. That RLS/PLM could be attributed to enhanced extracellular striatal dopamine but also occur in the setting of striatal dopamine loss is seemingly contradictory. This might be reconciled if one posits that primary/idiopathic RLS/PLM is rooted in a fundamental inefficiency of striatal dopaminergic signaling at post- rather than presynaptic sites. This would account for observations that the prevalence of RLS in PD patients lacking intact nigrostriatal dopaminergic axons mirrors that observed in the general population [53]. Alternatively, this could point to the principal pathophysiology residing outside dopaminergic nigrostriatal pathways, for example, in dopaminergic diencephalospinal pathways. As these pathways terminate largely in the dorsal horn where they likely inhibit superficial and deep tissue afferents, their dysfunction would provide the most parsimonious, unifying explanation for augmentation of the FR observed in RLS and the occurrence of abnormal sensations. Only one very preliminary experimental study has tested whether dopaminergic diencephalospinal pathway lesions can induce RLS/PLM [54]. While the findings are suggestive, they lack adequate behavioral analysis and a comprehensive accounting of the synaptic, cellular, and network mechanisms responsible.

The precise neural pathways that could convey the consequences of impaired striatal dopamine neurotransmission to spinal circuits that ultimately generate RLS/PLM begin with the striato-pallidal or striato-nigral pathways, as first suggested by Askenasy [55]. Pathways exiting the striatum ultimately engage the internal segment of the globus pallidus (GPi) and substantia nigra pars reticulata (SNr), which in turn concentrate their outputs upon thalamocortical networks, the deep layers of the superior colliculus, and the brainstem pedunculopontine region [56]. Underactivity of D₂ receptor-responsive sensorimotor striatal circuits would be expected to enhance GPi and SNr inhibitory influences to each of these targets [57]. Amelioration of RLS symptoms following pallidotomy in a patient with coexistent, idiopathic PD provides direct evidence, albeit anecdotal, that activity in basal ganglia sensorimotor circuits modulates RLS expression [58]. Our ongoing experience remains consistent in observing near elimination of PLM by pallidotomy in some PD patients [56]. Removal of pathologic pallidal output to the upper brainstem and, in turn, of descending influences upon ponto-medullary inhibition of spinal motor circuits, is the most parsimonious explanation for these benefits (namely, reversal of spinal disinhibition). Further confirmation and delineation of this multisynaptic pathway linking the basal ganglia and spinal cord is needed, but it is a plausible construct which accounts for several findings in RLS/PLM including: (1) enhancement in excitability and impaired habituation of brainstem-mediated aspects of the blink reflex [43,44]; (2) activation of synaptic or cellular elements in the mesopontine reticular formation visualized by functional magnetic resonance imaging [59]; and (3) lesion load below the tentorium (e.g. in the brainstem) correlating best with PLM in multiple sclerosis [60].

The considerations above do not preclude the fact that PLM can be modified by other central pathways. In fact, rather than exhibiting a unique pathology, it is reasonable to posit that PLM can arise from one of several central sources. From the perspective of spinal reflexes and local motor circuits, factors favoring PLM expression can be categorized into two groups: the first reflecting disinhibition due to interruption of descending inhibitory pathways, and a second including facilitation due to direct enhancement of neural activity. An example of the former includes modulation by the pyramidal motor system, given reports that patients with hemiplegia due to cerebrovascular disease exhibit PLM that predominate in the hemiplegic limbs [61]. Incidental damage to corticospinal fibers in nonhuman primates also unveils periodic movements that occur synchronously in the contralateral upper and lower limbs during non-REM sleep (personal observations). Enhancement of neural excitability by monoamines, on the other hand, likely accounts for the widely accepted clinical experience that RLS/PLM can be exacerbated by a wide array of antidepressants, including those designed to selectively increase synaptic availability of serotonin (namely, serotonin reuptake inhibitors) [62,63]. Even in a majority of normal volunteers lacking pre-existing RLS/PLM, the norepinephrine and serotonin reuptake blocker venlafaxine (Effexor[®]) induces PLM to greater than 25/hour of sleep [64]. It is unknown which specific neural substrates mediate these observed effects, given the ubiquitous nature of monoaminergic spinal innervation outlined above. Plausible loci are multifold and include direct enhancement of motoneuron responsiveness, 5-HT₂ receptor-mediated facilitation of the FR [65], and medullary raphe-mediated enhancement of spinal nociceptive transmission [66]. Decrements in monoaminergic neural integrity with aging, particularly of dopaminergic nigrostriatal cells, may also figure prominently in the pathophysiology of PLM given their steadily increasing prevalence along the lifespan. Thus, redundancy intrinsic to sensorimotor networks and the general lack of 'command' neurons (i.e. neurons both necessary and sufficient to generate a normal or pathologic motor behavior such as PLM) could readily account for heterogeneity as it regards the etiology of RLS/PLM and also account for variability in treatment responses.

2.2. PLM in Parkinson's disease (PD) and animal models of PD

Disordered sleep in the form of excessive nocturnal movement in PD was first noted in Parkinson's nineteenth-century Essay on the Shaking Palsy, and these findings have subsequently been confirmed and extended [67]. Periodic limb movements both with and without RLS complaints are common [53], occurring at a greater prevalence than in conditions lacking severe nigrostriatal dopamine neuron loss such as aging, Alzheimer's disease [51] and multisystem atrophy [52]. The intrinsic pathology in PD itself is likely to be the major contributor to these findings because early and/or untreated PD patients clearly exhibit disturbed sleep that includes excessive nocturnal movement [68–71]. Moreover, rats [50] and non-human primates [72] depleted of striatal dopamine bilaterally exhibit excessive nocturnal movement, and sleep in the parkinsonian patient deteriorates with disease progression [73-75]. Clinical improvements of PD seen with surgical interventions that restore balance to the altered basal ganglia neurotransmission accompanying nigrostriatal dopamine loss also improve sleep architecture [56,76]. Relative preservation of the dopaminergic diencephalospinal system in PD [77-79] makes it an unlikely contributor to the worsening of nocturnal movement in PD. The precise substrates accounting for the detrimental effects of nigrostriatal dopamine loss upon nocturnal movement are ill-defined. It is tempting to speculate that dopamine modulates brainstem circuits affecting PLM and REM sleep atonia. This does not occur via direct dopaminergic innervation of the brainstem but rather by indirect, multisynaptic routes linking the basal ganglia output nuclei with pontomedullary reticulospinal pathways via the dorsolateral pons, including the subcoeruleal region [56].

2.3. Sleepiness in PD: causes and potential neural substrates

Sleepiness in PD is common and very real. By self-report, approximately 10–50% of PD patients experience sleepiness. Many patients (nearly one half of those complaining of sleepiness) satisfy physiological criteria for narcolepsy, which is the prototypical disorder of sudden-onset sleep

[80,81]. Clinical and experimental experience concur that the disease itself, and not simply medication or sleep disruption, accounts for much of the attendant sleepiness. Dopamine loss within mesolimbic and mesocortical circuits, which is characteristic of patients with advanced disease, cognitive impairment, and drug-induced psychosis, appears most responsible [81]. Reports of sleepiness with dopaminomimetics are numerous and appear to be dose-related, but unrelated to specific class of agent (e.g. levodopa versus ergot and non-ergot-derived D2-like agonists), consistent with findings seen in healthy controls. These reports are largely anecdotal and have not documented or investigated the nature of related sleep-architecture changes. An excessive 14 h of total sleep, including excessive REM sleep, with pergolide use versus a normal 8.2 h of sleep without pergolide was recently reported [82]. Improvement in most measures of sleep continuity and architecture with pergolide was also observed. These findings suggest that drug-induced sleepiness in PD reflects an increased sleep drive versus an inability to remain awake. While only briefly alluded to in this report, another potential adverse effect of dopamine agonist use is enhancement of REM sleep-expression particularly the phasic components of REM sleep (including movement) - which are likely the physiological correlates of dream-like, hallucinatory behavior [83]. While some investigators suggest that obstructive sleep apnea (OSA) and PLM coexisting with PD may also contribute to sleepiness and intrusion of REM sleep into daytime naps (SOREMs), this is controversial since sleepiness in nonparkinsonian patients is not completely explained by the severity of OSA and PLM. Thus, premorbid sleepiness level, which is increasingly recognized to be a heritable trait in the general population, may also be a relevant factor in PD.

Dopamine pathways help to maintain sleep/wake homeostasis and are responsible for the wake-promoting effects of psychostimulants [84,85]. Thus, the parkinsonian state itself is a major factor in the expression of sleepiness and SOREMs. The point is best made in animal models of disease that control for potentially confounding variables such as age, comorbid conditions, and medications. Rats spend less of their subjective day awake following destruction of nigrostriatal pathways with bilateral, intrastriatal infusions of the dopamine toxin 6-hydroxydopamine [50]. Similarly, daytime sleepiness and SOREMs have been precipitated in a single non-human primate following systemic delivery of the dopamine neurotoxin 1-methyl,4phenyl-1,2,3,6-tetrahydropyridine (MPTP) [72]. The cellular and subcellular substrates underlying these diseaserelated effects remain ill-defined. These phenomena may reflect loss of dopamine's effects upon neural excitability in any one of a number of nuclei, which if unaffected by disease might otherwise serve to maintain normal states of thalamocortical excitability. One plausible substrate is novel dopamine pathways that simultaneously target movement-related subcortical circuits and the thalamus, which is

the final station for modulating cortical arousal [86]. Thalamocortical neuron firing rates and patterns are altered in non-human primates by focal applications of various dopaminomimetics [87].

Another plausible substrate, given the narcolepsy-like phenotype seen in many 'sleepy' PD patients, is hypocretin-containing neurons in the lateral hypothalamus known to degenerate in primary narcolepsy/cataplexy [88]. However, the findings of normal CSF hypocretin-1 levels in three PD patients with documented sleepiness argues against this hypothesis [89]. Other plausible neural substrates that deserve future investigation include targets of ventral tegmental area dopamine neurons including the prefrontal cortex, the cholinergic magnocellular basal forebrain, and midline thalamic nuclei. Alternatively, excessive daytime sleepiness (EDS) and SOREMs may reflect extranigral pathology in nuclei comprising the traditional ascending reticular activating system such as the dorsal raphe, locus coeruleus, and pedunculopontine tegmental nucleus (PPN). Dysregulation of the PPN region, an area known to promote thalamocortical arousal and REM sleep, may also be an important factor secondary to its position as a principal brainstem target of pathological basal ganglia outflow [56].

2.4. Summary

The preponderance of evidence argues that the pathophysiological basis of RLS/PLM resides in dopaminergic or dopamine-sensitive premotor circuits whose dysfunction results in disinhibition of spinal flexor reflex afferent pathways. Because many pathways converge upon this final common modulator of spinal sensorimotor function, the expression of RLS/PLM are likely influenced by numerous other supraspinal circuits, and are a common end point to numerous medical conditions/treatments. Two systems of primary interest in this regard include traditional nigrostriatal dopaminergic pathways and the less studied diencephalospinal dopaminergic pathways. The spectrum of sleepwake alterations seen with the loss of nigrostriatal dopamine characteristic of PD includes PLM and EDS, yet their precise pathological bases remain elusive. Until more data are forthcoming, the most prudent clinical and experimental approaches should proceed from the assumption that the parkinsonian condition represents an underlying diathesis to sleepiness and SOREM expression that can be exaggerated by numerous coexistent factors including use of dopamine agonists and levodopa, sedative-hypnotics, and potentially other medications; primary sleep disorders; and potentially comorbid conditions such as depression. While sleepiness observed with levodopa and dopamine agonists in PD has been increasingly publicized, these agents also cause sleepiness in drug-naïve controls. Thus, how often drug use in PD contributes further to sleepiness remains unresolved.

Acknowledgements

The authors' work is supported by PHS Grants NS-36697, NS-40221, and the Restless Legs Syndrome Foundation.

References

- Kuypers H. Anatomy of the descending pathways. In: Brookhart J, Mountcastle V, editors. Handbook of physiology – the nervous system, Bethesda, MD: American Physiological Society, 1981. pp. 597–666.
- [2] Holstege G. The somatic motor system. Prog Brain Res 1996;107:9– 26.
- [3] Rho M, Lavoie S, Drew T. Effects of red nucleus stimulation on the locomotor pattern and timing in the intact cat: a comparison with the motor cortex. J Neurophysiol 1999;81:2297–2315.
- [4] Cheney P, Fetz E, Mewes K. Neural mechanisms underlying corticospinal and rubrospinal control of limb movements. Prog Brain Res 1991;87:213–252.
- [5] Darian-Smith I, Burman K, Darian-Smith C. Parallel pathways mediating manual dexterity in the macaque. Exp Brain Res 1999;128:101–108.
- [6] Holstege JC, Kuypers HG. Brainstem projections to spinal motoneurons: an update. Neuroscience 1987;23:809–821.
- [7] Yates BJ, Smail JA, Stocker SD, Card JP. Transneuronal tracing of neural pathways controlling activity of diaphragm motoneurons in the ferret. Neuroscience 1999;90:1501–1513.
- [8] Billig I, Foris JM, Card JP, Yates BJ. Transneuronal tracing of neural pathways controlling an abdominal muscle, rectus abdominis, in the ferret. Brain Res 1999;820:31–44.
- [9] Hermann GE, Bresnahan JC, Holmes GM, Rogers RC, et al. Descending projections from the nucleus raphe obscurus to pudendal motoneurons in the male rat. J Comp Neurol 1998;397:458–474.
- [10] Holstege G, Bandler R, Saper CB. The emotional motor system. Prog Brain Res 1996;107:3–6.
- [11] Holstege JC. The ventro-medial medullary projections to spinal motoneurons: ultrastructure, transmitters and functional aspects. Prog Brain Res 1996;107:159–181.
- [12] Bjorklund A, Skagerberg G. Evidence for a major spinal cord projection from the diencephalic A11 dopamine cell group in the rat using transmitter-specific fluorescent retrograde tracing. Brain Res 1979;177:170–175.
- [13] Hokfelt T, Phillipson O, Goldstein M. Evidence for a dopaminergic pathway in the rat descending from the A11 cell group to the spinal cord. Acta Physiol Scand 1979;107:393–395.
- [14] Skagerberg G, Bjorklund A, Lindvall O, Schmidt RH. Origin and termination of the diencephalo-spinal dopamine system in the rat. Brain Res Bull 1982;9:237–244.
- [15] Skagerberg G, Lindvall O. Organization of diencephalic dopamine neurones projecting to the spinal cord in the rat. Brain Res 1985;342:340–351.
- [16] Nygren L-G, Olson L. A new major projection from locus coeruleus: the main source of noradrenergic nerve terminals in the ventral and dorsal columns of the spinal cord. Brain Res 1977;132:85–93.
- [17] Holets V, Hokfelt T, Rokaeus A, Terenius L, et al. Locus coeruleus neurons in the rat containing neuropeptide Y, tyrosine hydroxylase or galanin and their efferent projections to the spinal cord, cerebral cortex and hypothalamus. Neuroscience 1988;24:893–906.
- [18] Pickel V, Segal M, Bloom F. A radioautographic study of the efferent pathways of the nucleus locus coeruleus. J Comp Neurol 1974;155:15–42.
- [19] Westlund K, Bowker R, Ziegler M, Coulter J. Descending noradrenergic projections and their spinal terminations. Prog Brain Res 1982;57:219–238.
- [20] Carr PA, Pearson JC, Fyffe RE. Distribution of 5-hydroxytryptamine-

immunoreactive boutons on immunohistochemically-identified Renshaw cells in cat and rat lumbar spinal cord. Brain Res 1999;823:198–201.

- [21] Arvidsson U, Cullheim S, Ulfhake B, Bennett GW, et al. 5-Hydroxytryptamine, substance P, and thyrotropin-releasing hormone in the adult cat spinal cord segment L7: immunohistochemical and chemical studies. Synapse 1990;6:237–270.
- [22] Ellenberger H, Vera P, Feldman J, Holets V. Multiple putative neuromessenger inputs to the phrenic nucleus in rat. J Chem Neuroanat 1992;5:375–382.
- [23] Nicholas A, Pieribone V, Arvidsson U, Hokfelt T. Serotonin-, substance P- and glutamate/aspartate-like immunoreactivities in medullo-spinal pathways of rat and primate. Neuroscience 1992;48:545–549.
- [24] Fung SI, Chan JY, Manzoni D, White SR, et al. Cotransmittermediated locus coeruleus action on motoneurons. Brain Res Bull 1994;35:423–432.
- [25] Liu R, Fung S, Reddy V, Barnes C. Localization of glutamatergic neurons in the dorsolateral pontine tegmentum projecting to the spinal cord of the cat with a proposed role of glutamate on lumbar motoneuron activity. Neuroscience 1995;64:193–208.
- [26] Holstege JC, Van Dijken H, Buijs RM, Goedknegt H, et al. Distribution of dopamine immunoreactivity in the rat, cat and monkey spinal cord. J Comp Neurol 1996;376:631–652.
- [27] Magoun H, Rhines R. An inhibitory mechanism in the bulbar reticular formation. J Neurophysiol 1946;9:165–171.
- [28] Peterson B. Session chairman's overview: participation of pontomedullary reticular neurons in specific motor activity. In: Hobson J, Brazier M, editors. The reticular formation revisited: specifying function for a non-specific system, New York: Raven Press, 1980. pp. 171–192.
- [29] White SR, Fung SJ, Jackson DA, Imel KM. Serotonin, norepinephrine and associated neuropeptides: effects on somatic motoneuron excitability. Prog Brain Res 1996;107:183–199.
- [30] Smith DO, Lowe D, Temkin R, Jensen P, et al. Dopamine enhances glutamate-activated currents in spinal motoneurons. J Neurosci 1995;15:3905–3912.
- [31] Barbeau H, Rossignol S. Initiation and modulation of the locomotor pattern in the adult chronic spinal cat by noradrenergic, serotonergic and dopaminergic drugs. Brain Res 1991;546:250–260.
- [32] Kiehn O, Kjaerulff O. Spatiotemporal characteristics of 5-HT and dopamine-induced rhythmic hindlimb activity in the in vitro neonatal rat. J Neurophysiol 1996;75:1472–1482.
- [33] Katz P. Neurons, networks, and motor behavior. Neuron 1996;16:245–253.
- [34] Noga BR, Bras H, Jankowska E. Transmission from group II muscle afferents is depressed by stimulation of locus coeruleus/subcoeruleus, Kolliker-Fuse and raphe nuclei in the cat. Exp Brain Res 1992;88:502–516.
- [35] Jankowska E, Lackberg ZS, Dyrehag LE. Effects of monoamines on transmission from group II muscle afferents in sacral segments in the cat. Eur J Neurosci 1994;6:1058–1061.
- [36] Skoog B, Noga B. Dopaminergic control of transmission from group II muscle afferents to spinal neurones in the cat and guinea-pig. Exp Brain Res 1995;105:39–47.
- [37] Garraway SM, Hochman S. Comparison of synaptic and cellular actions of serotonin, noradrenaline, dopamine and acetylcholine in deep dorsal horn neurons. Soc Neurosci Abst 1999;25:922.
- [38] Freeman A, Ranadive V, Rye D. Human forebrain devoid of brainstem influences exhibits EEG and neuroendocrine rhythms [abstract]. Sleep 2000;23(Suppl. 2):A347–A348.
- [39] Yokota T, Hirose K, Tanabe H, Tsukagoshi H. Sleep-related periodic leg movements (nocturnal myoclonus) due to spinal cord lesion. J Neurol Sci 1991;104:13–18.
- [40] Dickel M, Renfrow S, Moore P, Berry R. Rapid eye movement sleep periodic leg movements in patients with spinal cord injury. Sleep 1994;17:733–738.

- [41] Lee M, Choi Y, Lee S. Sleep-related periodic leg movements associated with spinal cord lesions. Mov Disord 1996;11:719–722.
- [42] de Mello MT, Poyares DL, Tufik S. Treatment of periodic leg movements with a dopaminergic agonist in subjects with total spinal cord lesions. Spinal Cord 1999;37:634–637.
- [43] Wechsler L, Stakes J, Shahani B, Busis N. Periodic leg movements of sleep (nocturnal myoclonus): an electrophysiological study. Ann Neurol 1986;19:168–173.
- [44] Briellmann R, Rosler K, Hess C. Blink reflex excitability is abnormal in patients with periodic leg movements in sleep. Mov Disord 1996;11:710–714.
- [45] Bara-Jimenez W, Aksu M, Graham B, Sato S, et al. Periodic limb movements in sleep – state dependent excitability of the spinal flexor reflex. Neurology 2000;54:1609–1615.
- [46] Montplaisir J, Godbout R, Boghen D, DeChamplain J, et al. Familial restless legs with periodic movements in sleep: electrophysiologic, biochemical, and pharmacologic study. Neurology 1985;35:130–134.
- [47] Turjanski N, Lees A, Brooks D. Striatal dopaminergic function in restless legs syndrome. Neurology 1999;52:932–937.
- [48] Staedt J, Stoppe G, Kogler A, Riemann H, et al. Nocturnal myoclonus syndrome (periodic movements in sleep) related to central dopamine D2-receptor alteration. Eur Arch Psychiatr Clin Neurosci 1995;245:8–10.
- [49] Eisensehr I, Wetter TC, Linke R, Noachtar S, et al. Normal IPT and IBZM SPECT in drug-naive and levodopa-treated idiopathic restless legs syndrome. Neurology 2001;57:1307–1309.
- [50] Decker M, Keating G, Freeman A, Rye D. Parkinsonian-like sleepwake architecture in rats with bilateral striatal 6-OHDA lesions. Soc Neurosci Abstr 2000;26:1514.
- [51] Bliwise D, Rye D, Dihenia B, Yesavage JA, et al. Periodic leg movements in elderly patients with Parkinsonism [abstract]. Sleep 1998;21(Suppl.):196.
- [52] Wetter T, Collado-Seidel V, Pollmacher T, Yassouridis A, et al. Sleep and periodic leg movement patterns in drug-free patients with Parkinson's disease and multiple system atrophy. Sleep 2000;23:361–367.
- [53] Ondo WG, Vuong KV, Khan H, Atassi F, et al. Daytime sleepiness and other sleep disorders in Parkinson's disease. Neurology 2001;57:1392–1396.
- [54] Ondo WG, He Y, Rajasekaran S, Le WD. Clinical correlates of 6hydroxydopamine injections into A11 dopaminergic neurons in rats: a possible model for restless legs syndrome. Mov Disord 2000;15:154– 158.
- [55] Askenasy J, Weitzman E, Yahr M. Are periodic movements in sleep a basal ganglia dysfunction? J Neural Transm 1987;70:337–347.
- [56] Rye D. Contributions of the pedunculopontine region to normal and altered REM sleep. Sleep 1997;20:757–788.
- [57] Wichmann T, DeLong MR. Functional and pathophysiological models of the basal ganglia. Curr Opin Neurobiol 1996;6:751–758.
- [58] Rye D, DeLong M. Amelioration of sensory limb discomfort of restless legs syndrome by pallidotomy. Ann Neurol 1999;46:800–801.
- [59] Bucher S, Seelos K, Oertel W, Reiser M, et al. Cerebral generators involved in the pathogenesis of the restless legs syndrome. Ann Neurol 1997;4:639–645.
- [60] Ferini-Strambi L, Filippi M, Martinelli V, Oldani A, et al. Nocturnal sleep study in multiple sclerosis: correlations with clinical and brain magnetic resonance imaging findings. J Neurol Sci 1994;125:194– 197.
- [61] Inami Y, Shimizu T, Iijima S, Okawa M, et al. Nocturnal myoclonus syndrome in patients with hemiplegia due to cerebrovascular disease [abstract]. Sleep Res 1987;16:509.
- [62] Morgan J, Brown T, Wallace E. Monoamine oxidase inhibitors and sleep movements. Am J Psychiatry 1994;151:782–783.
- [63] Bakshi R. Fluoxetine and restless legs syndrome. J Neurol Sci 1996;142:151–152.
- [64] Salin-Pascual R, Galicia-Polo L, Drucker-Colin R. Sleep changes after 4 consecutive days of venlafaxine administration in normal volunteers. J Clin Psychiatry 1997;58:348–350.

- [65] Skarsfeldt T, Arnt J, Hyttel J. L-5-HTP facilitates the electrically stimulated flexor reflex in pitched rats: evidence for 5-HTP2-receptor mediation. Eur J Pharmacol 1990;176:135–142.
- [66] Zhuo M, Gebhart G. Biphasic modulation of spinal nociceptive transmission from the medullary raphe nuclei in the rat. J Neurophysiol 1997;78:746–758.
- [67] Rye D, Bliwise D. Movement disorders specific to sleep and the nocturnal manifestations of waking movement disorders. In: Watts R, Koller W, editors. Movement disorders: neurologic principles and practice, New York: McGraw-Hill, Inc, 1997. pp. 687–713.
- [68] Kales A, Ansel R, Markham C, Scharf MB, et al. Sleep in patients with Parkinson's disease and normal subjects prior to and following levodopa administration. Clin Pharmacol Ther 1971;12:397–406.
- [69] Bergonzi P, Chiurulla C, Cianchetti C, Tempesta E. Clinical pharmacology as an approach to the study of biochemical sleep mechanisms: the action of L-dopa. Confin Neurol 1974;36:5–22.
- [70] Bergonzi P, Chiurulla C, Gambi D, Mennuni G, et al. L-dopa plus dopadecarboxylase inhibitor. Sleep organization in Parkinson's syndrome before and after treatment. Acta Neurol Belg 1975;75:5–10.
- [71] Rye D, Johnston L, Watts R, Bliwise D. Juvenile Parkinson's disease with REM behavior disorder, sleepiness and daytime REM-onsets. Neurology 1999;53:1868–1870.
- [72] Daley J, Turner R, Bliwise D, Rye D. Nocturnal sleep and daytime alertness in the MPTP-treated primate [abstract]. Sleep 1999;22(Suppl.):S218–S219.
- [73] Friedman A. Sleep pattern in Parkinson's disease. Acta Med Pol 1980;21:193–199.
- [74] Schneider E, Ziegler B, Maxion H, Diehl R, Tempesta E, et al. Sleep in parkinsonian patients under levodopa. Results of a long-term follow-up study. 3rd European Congress Sleep Research. Basel: Karger, 1976. pp. 447–450.
- [75] Emser W, Hoffmann K, Stolz T. Sleep disorders in diseases of the basal ganglia. Interdisciplinary topics in gerontology, Basel: Karger, 1987. pp. 144–157.
- [76] Arnulf I, Bejjani B, Garma L, Bonnet AM, et al. Improvement of sleep architecture in PD with subthalamic nucleus stimulation. Neurology 2000;55:1732–1734.
- [77] Matzuk M, Saper C. Preservation of hypothalamic dopaminergic neurons in Parkinson's disease. Ann Neurol 1985;18:552–555.

- [78] Scatton B, Dennis T, L'Heureux R, Monfort J-C, et al. Degeneration of noradrenergic and serotonergic but not dopaminergic neurones in the lumbar spinal cord of parkinsonian patients. Brain Res 1986;380:181–185.
- [79] Sofic E, Riederer P, Gsell W, Gavranovic M, et al. Biogenic amines and metabolites in spinal cord of patients with Parkinson's disease and amyotrophic lateral sclerosis. J Neural Transm Park Dis Dement Sect 1991;3:133–142.
- [80] Rye DB, Bliwise DL, Dihenia B, Gurecki P. FAST TRACK: daytime sleepiness in Parkinson's disease. J Sleep Res 2000;9:63–69.
- [81] Rye DB, Jankovic J. Emerging views of dopamine in modulating sleep/wake state from an unlikely source: PD. Neurology 2002;58:341–346.
- [82] Ulivelli M, Rossi S, Lombard C, Bartalini S, et al. Polysomnographic characterization of pergolide-induced 'sleep attacks' in an idiopathic PD patient. Neurology 2002;58:462–465.
- [83] Arnulf I, Bonnet A, Damier P, Beijani BP, et al. Hallucinations, REM sleep, and Parkinson's disease: a medical hypothesis. Neurology 2000;55:281–288.
- [84] Kanabayashi T, Honda K, Kodama T, Mignot E, et al. Implication of doapminergic mechanisms in the wake-promoting effects of amphetamine: a study of D- and L-derivatives in canine narcolepsy. Neuroscience 2000;99:651–659.
- [85] Wisor J, Nishino S, Sora I, Uhl G, et al. Dopaminergic role in stimulant-induced wakefulness. J Neurosci 2001;21:1787–1794.
- [86] Freeman A, Ciliax B, Bakay R, Daley J, et al. Nigrostriatal collaterals to thalamus degenerate in parkinsonian animal models. Ann Neurol 2001;50:321–329.
- [87] Rye DB, Daley JT, Freeman AA, Bliwise DL. Daytime sleepiness in idiopathic Parkinson's disease. In: Bedard MA, Agid Y, Chouinard S, Fahn S, et al., editors. Mental and behavioral dysfunction in movement disorders. Humana Press: Totowa, NJ, 2002 (in press).
- [88] Silber MH, Rye DB. Solving the mysteries of narcolepsy. Neurology 2001;56:1616–1618.
- [89] Overeem S, van Hilten JJ, Ripley B, Mignot E, et al. Normal hypocretin-1 levels in Parkinson's disease patients with excessive daytime sleepiness. Neurology 2002;58:465–468.